

Institut für Lebensmittelsicherheit und -hygiene  
der Vetsuisse-Fakultät Universität Zürich  
Direktor: Prof. Dr. med. vet. Dr. h.c. Roger Stephan

Arbeit unter wissenschaftlicher Betreuung von  
Prof. Dr. med. vet. Dr. h. c. Roger Stephan

***Listeria monocytogenes* in Surface Waters in Switzerland:  
Identification of Epidemic Clones and Characterization of  
Virulence Traits**

**Inaugural-Dissertation**

zur Erlangung der Doktorwürde der  
Vetsuisse-Fakultät Universität Zürich

vorgelegt von

**Susanne Raschle**

Tierärztin  
von Mosnang, St. Gallen

genehmigt auf Antrag von  
Prof. Dr. med. vet. Dr. h. c. Roger Stephan, Referent

**2021**



# Inhaltsverzeichnis

1 Summary	4
2 Zusammenfassung	5
3 Printversion Publikation scientific reports	6
4 Anhänge	18
Danksagung/Acknowledgement	
Curriculum Vitae	

# 1 Summary

*Listeria monocytogenes* is an opportunistic pathogen that is widely distributed in the environment. The aquatic environment may represent a potential source for the transmission of *L. monocytogenes* to animals and the food chain. The present study assessed the occurrence of *L. monocytogenes* in surface water samples from rivers and streams throughout Switzerland. Whole genome sequence (WGS) data of the isolates were used to assign them to lineages, serotypes, sequence types (STs), and clonal complexes (CCs), and to assess virulence genotypes. Twenty-five (13%) of the surface water samples were positive for the presence of *L. monocytogenes*. The isolates belonged to major lineages I and II, with the majority assigned to either serotype 1/2b (48%), or 4b (44%). The predominant CCs identified were CC1 (20%), CC4 (16%), and CC412 (16%), all of which are implicated in listeriosis outbreaks and sporadic cases of human and animal infection worldwide. Two (8%) of the isolates belonged to CC6 which is an emerging hypervirulent clone. All isolates contained intact genes associated with invasion and infection, including *inlA/B* and *prfA*. The four CC4 isolates all harbored *Listeria* pathogenicity island 4 (LIPI-4), which confers hypervirulence. The occurrence of *L. monocytogenes* in river ecosystems may contribute to the dissemination and introduction of clinically highly relevant strains to the food chain.

**Key words:** *Listeria monocytogenes*, aquatic environment, clones

## 2 Zusammenfassung

*Listeria monocytogenes* ist ein opportunistischer pathogener Erreger und in der Umwelt weit verbreitet. Die Übertragung von *L. monocytogenes* auf Tiere und die Lebensmittelkette erfolgt möglicherweise auch über Gewässer. In dieser Studie wurden Proben aus Flüssen und Bächen, die schweizweit erhoben wurden, auf das Vorkommen von *L. monocytogenes* untersucht. Gesamtgenomdaten wurden verwendet, um die Isolate in Linien, Serotypen, Sequenztypen (STs) und klonale Komplexe (CCs) einzuteilen und die Virulenzgen Muster zu bestimmen. Fünfundzwanzig (13%) der Wasserproben waren *L. monocytogenes* positiv. Die Isolate gehörten zu den Hauptlinien I und II, wobei die Mehrheit entweder dem Serotyp 1/2b (48 %) oder 4b (44 %) zugeordnet werden konnte. Die Isolate gehörten mehrheitlich zu CC1 (20 %), CC4 (16 %) und CC412 (16 %). Zwei (8 %) der Isolate gehörten zum CC6, einem hypervirulenten clonalen Komplex. Alle Isolate enthielten intakte Virulenzgene, einschliesslich *inlA/B* und *prfA*. Die vier CC4-Isolate enthielten alle zudem die Pathogenitätsinsel 4 (LIPI-4). Das Vorkommen von *L. monocytogenes* in Fliessgewässern könnte zur Entstehung und Verbreitung klinisch hoch relevanter Stämme auch die Lebensmittelkette beitragen.

**Schlüsselwörter:** *Listeria monocytogenes*, Fliessgewässer, Gesamtgenomdaten

### **3 Printversion Publikation scientific reports**

Environmental dissemination of pathogenic *Listeria monocytogenes* in flowing surface waters in Switzerland



OPEN

# Environmental dissemination of pathogenic *Listeria monocytogenes* in flowing surface waters in Switzerland

Susanne Raschle, Roger Stephan, Marc J. A. Stevens, Nicole Cernela, Katrin Zurfluh, Francis Muchaamba & Magdalena Nüesch-Inderbinen

*Listeria monocytogenes* is an opportunistic pathogen that is widely distributed in the environment. The aquatic environment may represent a potential source for the transmission of *L. monocytogenes* to animals and the food chain. The present study assessed the occurrence of *L. monocytogenes* in 191 surface water samples from rivers, streams and inland canals throughout Switzerland. Twenty-five (13%) of the surface water samples contained *L. monocytogenes*. Whole genome sequence (WGS) data were used to characterize the 25 isolates. The isolates belonged to major lineages I and II, with the majority assigned to either serotype 1/2a (48%), or 4b (44%). The predominant CCs identified were the hypervirulent serotype 4b clones CC1 and CC4, and the serotype CC412; all three have been implicated in listeriosis outbreaks and sporadic cases of human and animal infection worldwide. Two (8%) of the isolates belonged to CC6 which is an emerging hypervirulent clone. All isolates contained intact genes associated with invasion and infection, including *inlA/B* and *prfA*. The four CC4 isolates all harbored *Listeria* pathogenicity island 4 (LIPI-4), which confers hypervirulence. The occurrence of *L. monocytogenes* in river ecosystems may contribute to the dissemination and introduction of clinically highly relevant strains to the food chain.

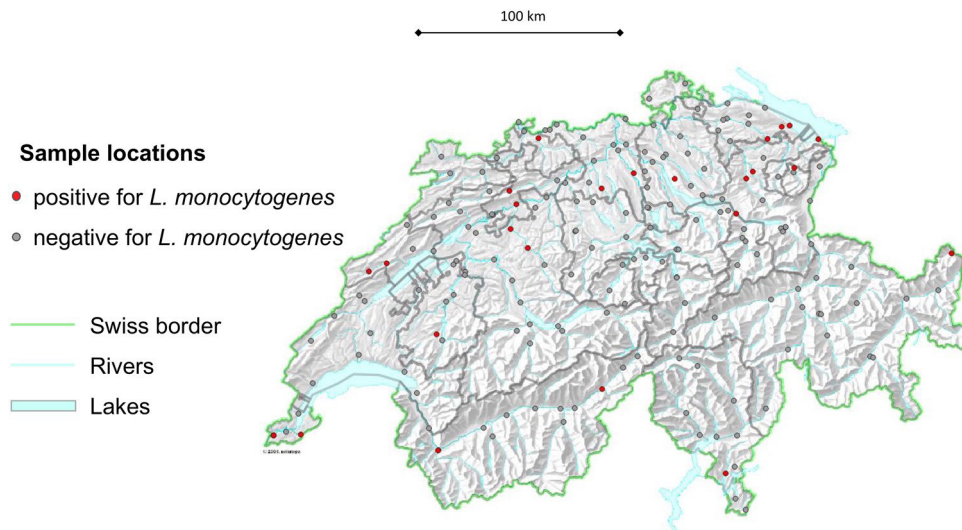
*Listeria monocytogenes* is a ubiquitous Gram-positive bacterium and an opportunistic pathogen that causes listeriosis in humans and in some animal species<sup>1</sup>. In humans, listeriosis is a potentially lethal infection, with the vulnerable populations such as the elderly, pregnant women, and immunocompromised persons at particular risk for meningitis, sepsis, premature birth, or abortion<sup>2</sup>. In animals, particularly in ruminants, listeriosis can manifest as rhombencephalitis and in its septicemic form may cause abortion, stillbirth, and death<sup>3</sup>.

*Listeria monocytogenes* is classified into four major evolutionary lineages and four PCR based serogroups<sup>4,5</sup>. The majority of *L. monocytogenes* isolates belong to lineage I that is associated with serotypes 1/2b, 3b and 4b, and to lineage II which includes serotype 1/2a, 1/2c, 3a, and 3c<sup>5</sup>. Further, multilocus sequence typing (MLSTs) subdivides these categories into numerous clonal complexes (CCs) and sequence types (STs). Certain serotypes specifically 1/2a, 1/2b and 4b, and certain CCs including hypervirulent strains assigned to CC1, CC4 and CC6, are frequently encountered in clinical cases<sup>5,6</sup>.

Following survival within the gastrointestinal (GI) tract, multiple virulence factors (VFs) enable *L. monocytogenes* to invade and survive within mammalian host cells<sup>7</sup>. Key VFs include InlA and InlB, which belong to a family of internalins that allow the bacteria to incorporate within vacuoles of the host cells, listeriolysin O (LLO) and phospholipases PlcA and PlcB which are involved in vacuolar lysis, as well as ActA, which provides motility within the cytosol<sup>1,8,9</sup>.

*Listeria monocytogenes* virulence genes are mostly organized in clusters located throughout the chromosome, such as the internalin gene operon, several listerial pathogenicity islands (LIPI-1 to LIPI-4), and stress survival islets (SSIs). *L. monocytogenes* harboring LIPI-4 are considered hypervirulent and are associated with enhanced invasion and neural and placental infection<sup>6</sup>. By contrast, *L. monocytogenes* isolated from food processing environments and from food are frequently associated with reduced pathogenicity due to truncated and non-functional major virulence factors such as *inlA/B*<sup>10,11</sup>. Such InlA truncations are partially accountable for hypovirulence in *L. monocytogenes* CC9 and CC121, which are major CCs associated with a food origin<sup>6</sup>.

Institute for Food Safety and Hygiene, Vetsuisse Faculty, University of Zurich, Zurich, Switzerland. ✉ email: magdalena.nuesch-inderbinen@uzh.ch



**Figure 1.** Map of Switzerland showing surface waters, sampling locations, and sites of strain isolation. The map was created using the open source geographic information system (GIS) software QGIS version 3.10 (<https://qgis.org>).

Consumption of food, notably ready-to-eat food, fresh raw produce, and animal-derived food products that are consumed raw is the primary route of exposure of humans to pathogenic *L. monocytogenes*<sup>12</sup>. The occurrence and persistence of *Listeria* in processing plants are frequently caused by environmental recontamination at plant or farm level<sup>13</sup>. At farm level, the route of *L. monocytogenes* infection in ruminants is understood to be contaminated silage and contamination of farm environments including cattle bedding and water troughs<sup>14</sup>. The introduction sources of *L. monocytogenes* to farm animals, and the pathways permitting pathogenic *L. monocytogenes* entering the food chain are currently not completely understood. *Listeria* is widely distributed in the natural environment, with soil representing a key niche for the persistence of globally distributed *L. monocytogenes* isolates<sup>13</sup>. Soil runoff may contaminate water sources which then serve as reservoirs that transfer *Listeria* through the environment<sup>15</sup>. In addition, contaminated sewage or wastewater effluent is reportedly an important cause of *Listeria* in rivers<sup>16,17</sup>, with *L. monocytogenes* found more commonly than other *Listeria* species in surface waters in urban environments<sup>18</sup>. The aquatic ecosystem therefore provides an ideal setting for the circulation of *L. monocytogenes* between the habitats such as soil, plants, animals, natural and urban environments<sup>19</sup>. A number of studies have shown that *Listeria* occurs in waterways in farm environments and agricultural areas, including water which could be used for irrigation<sup>20,21</sup>. Several studies have reported the prevalence of *L. monocytogenes* in river water in the US and Canada, with serogrouping and pulsed field gel electrophoresis (PFGE) analysis revealing serogroups and pulsotypes that were similar to human *L. monocytogenes* isolates<sup>16,21</sup>. Furthermore, surface water strains may carry functional *inlA* genes thus being potentially virulent<sup>15,17</sup>. Although it is therefore recognized that pathogenic *L. monocytogenes* occur in surface water, whole genome sequencing-based information regarding the molecular diversity of *L. monocytogenes* occurring within the aquatic environment is currently limited.

This study was designed to evaluate the occurrence of *L. monocytogenes* in flowing surface waters throughout Switzerland and to characterize the isolated strains using whole genome analyses. We aimed to identify in the aquatic environment, any epidemic clones associated with human and animal infections in order to assess the relevance of such strains to public health as well as the health of animals. Further emphasis was placed on identifying virulence factors (VFs).

## Results

**Occurrence of *L. monocytogenes* in flowing surface water.** A total of 191 water samples were collected and analyzed. The geographical distribution of the 191 sampling sites is depicted in Fig. 1. Of the 190 sites, 141 (74%) were located downstream of a wastewater treatment plant (WWTP) (see Supplementary Table S1).

After enrichment, 25 (13%) samples revealed one to three presumptive colonies on OCLA plates. From each plate, one colony was selected for species identification and further analysis using whole genome sequencing (WGS). The positive samples were from sites situated between 280 and 1560 m above sea level, and 19 (76%) were located downstream of WWTPs (Table 1).

**Assignment of *L. monocytogenes* lineages and serotypes.** After implementation of WGS for the 25 isolates, in silico analyses including serotyping identified thirteen (52%) strains belonging to lineage I, including 11 strains representing serotype 4b, and two belonging to serotype 1/2b (Table 2). The remaining 12 (48%) strains belonged to lineage II and serotype 1/2a (Table 2). The general features of the 25 *Listeria monocytogenes* draft genomes are listed in Table 3.



Water sample ID	Sampling date (dd.mm.yy)	Location (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Downstream of WWTP
L_12	16.01.20	47° 19' 42" 8° 36' 54"	Fair (−2 to 8 °C)	580	Stream	+
L_28	27.01.20	47° 30' 56" 7° 43' 14"	Cloudy (1–3 °C)	290	River	+
L_36	27.01.20	47° 16' 49" 7° 31' 41"	Cloudy (1–3 °C)	680	Stream	+
L_41	11.02.20	47° 33' 6" 9° 19' 34"	Heavy rain (−1 to 5 °C)	420	Stream	+
L_42	11.02.20	47° 29' 56" 9° 13' 51"	Heavy rain (−1 to 5 °C)	470	River	+
L_44	11.02.20	47° 19' 25" 9° 5' 6"	Heavy rain (−1 to 5 °C)	600	Stream	−
L_49	11.02.20	47° 29' 31" 9° 33' 56"	Heavy rain (−1 to 5 °C)	400	River	+
L_50	11.02.20	47° 21' 16' 9° 7' 45"	Heavy rain (−1 to 5 °C)	630	River	+
L_51	11.02.20	47° 22' 0' 9° 24' 9"	Heavy rain (−1 to 5 °C)	800	Stream	+
L_52	11.02.20	47° 33' 23" 9° 22' 42"	Heavy rain (−1 to 5 °C)	400	River	+
L_57	17.02.20	47° 6' 35" 7° 32' 8"	Cloudy (6–12 °C)	480	Stream	+
L_58	17.02.20	47° 1' 28" 7° 38' 52"	Cloudy (6–12 °C)	560	River	+
L_72	24.02.20	47° 17' 21" 8° 8' 1"	Cloudy (12–15 °C)	490	Stream	+
L_86	02.03.20	47° 13' 14" 7° 34' 25"	Cloudy (2–8 °C)	420	River	+
L105	09.03.20	47° 21' 21" 8° 20' 45"	Rain (4–7 °C)	370	River	+
L111	09.03.20	47° 10' 3" 9° 0' 51"	Rain (4–7 °C)	430	Inland canal	+
L124	15.03.20	46° 57' 43" 10° 24' 53"	Fair (2–13 °C)	1560	Stream	−
L127	16.03.20	46° 57' 14" 6° 43' 35"	Fair (−1 to 10 °C)	730	Stream	+
L128	16.03.20	46° 54' 60" 6° 36' 39"	Fair (−1 to 10 °C)	730	River	+
L137	16.03.20	46° 10' 57" 6° 0' 36"	Fair (−1 to 16 °C)	350	River	−
L138	16.03.20	46° 10' 59" 6° 11' 1"	Fair (−1 to 16 °C)	400	Stream	−
L164	03.05.20	46° 0' 23" 8° 54' 48"	Fair (6–20 °C)	280	River	+
L174	12.05.20	46° 38' 14" 7° 3' 20"	Light rain (3–6 °C)	730	Stream	−
L180	12.05.20	46° 7' 7" 7° 4' 5"	Light rain (3–6 °C)	460	River	−
L188	12.05.20	46° 23' 30" 8° 7' 32"	Cloudy (5–13 °C)	990	River	+

**Table 1.** Key features of 25 surface water sampling sites testing positive for *Listeria monocytogenes*. DMS, degrees, minutes, and seconds; WWTP, Wastewater treatment plant.

**Identification of epidemic and outbreak clones.** To assess the clinical relevance of *L. monocytogenes* occurring in the aquatic environment, the genomes of the 25 strains were subjected to detailed in silico analysis. Based on the seven-gene MLST scheme provided by the BIGSdb-*L. monocytogenes* platform (<https://bigsdb.pasteur.fr/listeria>), a total of 11 STs which corresponded to 11 CCs were identified (Table 2). The predominant CCs were CC1 (20%), CC4 (16%), and CC412 (16%) (Table 2). *L. monocytogenes* serotype 4b CC1 and CC4 are epidemic clones associated with human listeriosis outbreaks worldwide, and both clones are also frequent among strains causing listeriosis in ruminants<sup>6,22,23</sup>. By contrast, *L. monocytogenes* serotype 1/2a CC412 is only sporadically associated with human infections but is prevalent among strains causing rhombencephalitis in cattle<sup>6,22</sup>. Two further strains (8%) were assigned to CC6, which is an emerging epidemic *L. monocytogenes* serotype 4b clone causing major outbreaks and severe forms of human listerial meningitis worldwide<sup>6,24</sup>. Other CCs identified among the strains in this study included *L. monocytogenes* serotype 1/2a CC7, CC11 and CC29, all of which have been associated with outbreaks occurring in the US between 1987 and 2011, and further found to represent environmental strains that persist within food production and livestock environments globally<sup>23,25–27</sup>. On the other hand, *L. monocytogenes* serotype 1/2a CC37 found in two samples in this study, is associated with abortion in small ruminants and cattle and found frequently in wildlife and ruminant environments<sup>13,25,28</sup>. Further, *L. monocytogenes* CC37 is prominent in milk samples from US dairy farms<sup>29</sup>.

**Analysis of strain relatedness.** The 25 *L. monocytogenes* strains belonged to 25 different cgMLST types (CTs), as shown in Table 2. The population structure of the strains was visualized by constructing a phylogenetic tree based on cgMLST. The isolates grouped according to lineages and serotypes, but were phylogenetically clearly distinct from each other, with  $\geq 10$  different alleles between each pair of neighboring isolates (Fig. 2). The genomes of the CC1, CC4, CC6, CC7, CC37, CC59, and CC412 isolates were compared with the available genomes of corresponding CCs present in the database of the Swiss National Reference Centre for Enteropathogenic Bacteria and *Listeria* (NENT) which collects all *L. monocytogenes* strains from confirmed human listeriosis cases nationwide and performs Illumina-based whole genome sequencing. The cgMLST-based phylogenetic trees are shown in Fig. 3 and the number of *L. monocytogenes* genomes in the NENT database are listed in Table 3. None of the 25 strains from this study clustered with a strain in the database, thereby ruling out a direct match with any *L. monocytogenes* reported from a case of human disease in Switzerland.

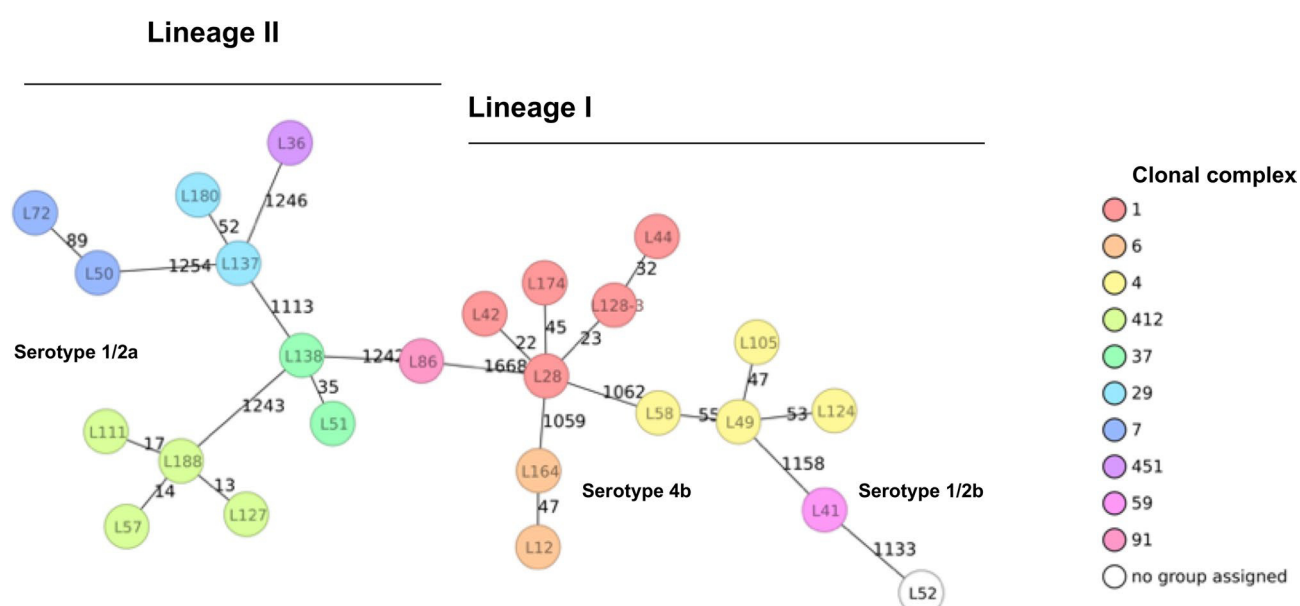
Strain ID	Location of isolation (DMS)	Date of isolation (dd.mm.yy)	Lineage	Serotype	CC	ST	cgMLST CT	inlA/B	LIPI-1	LIPI-3	LIPI-4	SSI-1	GenBank accession no
L28	47° 30' 56", 7° 43' 14"	27.01.20	I	4b	1	1	14,372	+	+	+	–	–	JACOFA000000000
L44	47° 19' 25", 9° 5' 6"	11.02.20	I	4b	1	1	14,376	+	+	+	–	–	JACOEW000000000
L42	47° 29' 56", 9° 13' 51"	11.02.20	I	4b	1	1	14,361	+	+	+	–	–	JACOEX000000000
L128-3	46° 54' 60", 6° 36' 39"	16.03.20	I	4b	1	1	14,359	+	+	+	–	–	JACOFH000000000
L174	46° 38' 14", 7° 3' 20"	12.05.20	I	4b	1	1	14,370	+	+	+	–	–	JACOFD000000000
L49	47° 29' 31", 9° 33' 56"	11.02.20	I	4b	4	4	14,365	+	+	+	+	–	JACOEV000000000
L58	47° 1' 28", 7° 38' 52"	17.02.20	I	4b	4	4	14,362	+	+	+	+	–	JACOEQ000000000
L105	47° 21' 21", 8° 20' 45"	09.03.20	I	4b	4	4	14,364	+	+	+	+	–	JACOFM000000000
L124	46° 57' 43", 10° 24' 53"	15.03.20	I	4b	4	4	14,366	+	+	+	+	–	JACOFJ000000000
L12	47° 19' 42", 8° 36' 54"	16.01.20	I	4b	6	6	14,375	+	+	+	–	–	JACOFK000000000
L164	46° 0' 23", 8° 54' 48"	03.05.20	I	4b	6	6	14,377	+	+	+	–	–	JACOFE000000000
L52	47° 33' 23", 9° 22' 42"	11.02.20	I	1/2b	224	2332	14,315	+	+	+	–	+	JACOE000000000
L41	47° 33' 6", 9° 19' 34"	11.02.20	I	1/2b	59	59	14,363	+	+	–	–	–	JACOEY000000000
L50	47° 21' 16", 9° 7' 45"	11.02.20	II	1/2a	7	7	14,367	+	+	–	–	+	JACOEU000000000
L72	47° 17' 21", 8° 8' 1"	24.02.20	II	1/2a	7	7	14,378	+	+	–	–	+	JACOEP000000000
L36	47° 16' 49", 7° 31' 41"	27.01.20	II	1/2a	11	451	14,371	+	+	–	–	–	JACOEZ000000000
L86	47° 13' 14", 7° 34' 25"	02.03.20	II	1/2a	14	91	14,314	+	+	–	–	–	JACOE000000000
L137	46° 10' 57", 6° 0' 36"	16.03.20	II	1/2a	29	29	14,368	+	+	–	–	–	JACOFG000000000
L180	46° 7' 7", 7° 4' 5"	12.05.20	II	1/2a	29	29	8916	+	+	–	–	–	JACOF000000000
L51	47° 22' 0', 9° 24' 9"	11.02.20	II	1/2a	37	37	14,374	+	+	–	–	–	JACOET000000000
L138	46° 10' 59", 6° 11' 1"	16.03.20	II	1/2a	37	37	7538	+	+	–	–	–	JACOFF000000000
L57	47° 6' 35", 7° 32' 8"	17.02.20	II	1/2a	412	412	14,369	+	+	–	–	–	JACOER000000000
L111	47° 10' 3', 9° 0' 51"	09.03.20	II	1/2a	412	412	14,360	+	+	–	–	–	JACOF000000000
L127	46° 57' 14", 6° 43' 35"	16.03.20	II	1/2a	412	412	14,373	+	+	–	–	–	JACOFI000000000
L188	46° 23' 30", 8° 7' 32"	12.05.20	II	1/2a	412	412	13,675	+	+	–	–	–	JACOFB000000000
LL195 <sup>a</sup>	n.a	n.a	I	4b	1	1	24	+	+	+	–	–	HF558398
N2306 <sup>a</sup>	n.a	n.a	I	4b	4	4	2506	+	+	+	+	–	CP011004
EGD-e <sup>a</sup>	n.a	n.a	II	1/2a	9	35	1	+	+	–	–	+	NC003210
N1546 <sup>a</sup>	n.a	n.a	II	1/2a	8	8	3614	+	+	–	–	+	CP013724
N12-1273 <sup>a</sup>	n.a	n.a	II	1/2a	412	412	9302	+	+	–	–	–	QYFZ000000000

**Table 2.** Key features of 25 *Listeria monocytogenes* isolates from flowing surface water and of five reference strains. <sup>a</sup> reference strains; CC, clonal complex; CT, cluster type; cgMLST, core genome multilocus sequence type; inlA/B, full length internalin genes A and B; LIPI, *Listeria monocytogenes* pathogenicity island; n.a., not applicable; SSI, stress survival islet; ST, sequence type; +, presence of gene(s); –, absence of gene(s).

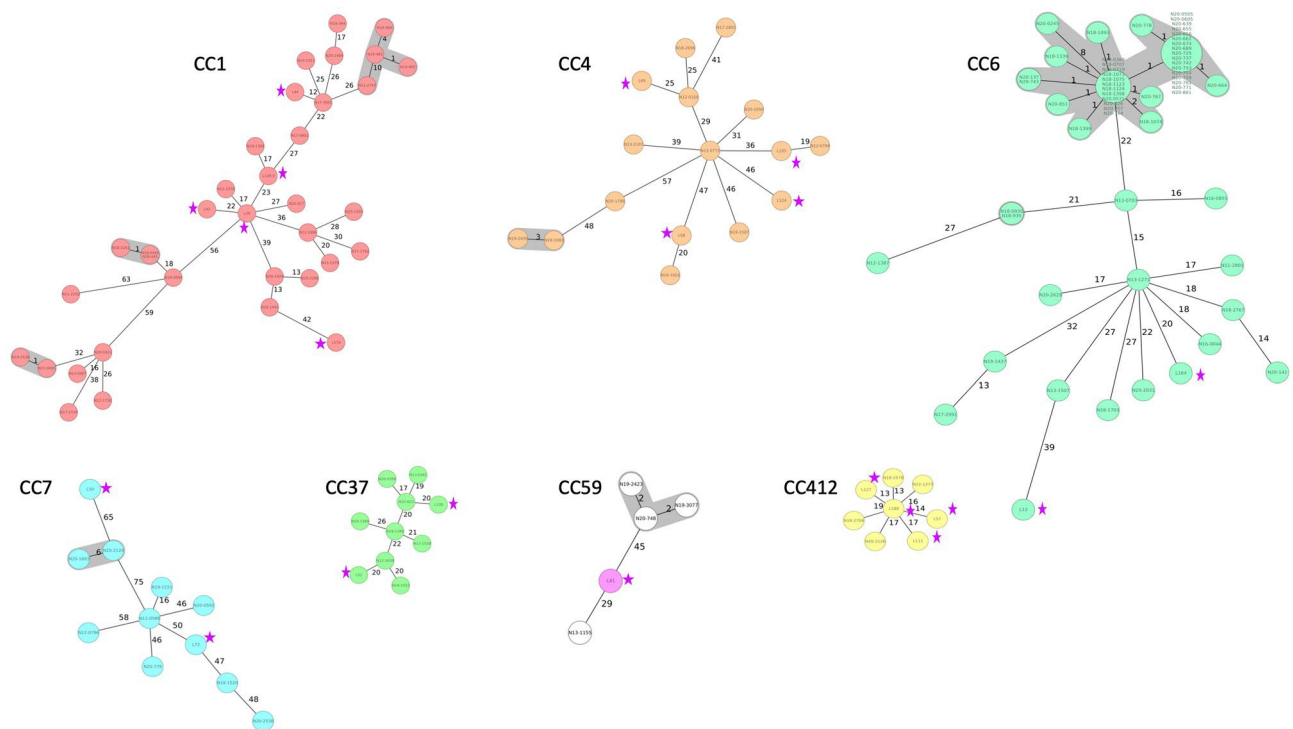
**Assessment of virulence attributes.** Pan-genomic analysis was performed to identify the presence of virulence genes associated with invasion of mammalian host cells, hypervirulence, or neural and placental infections.

Strain ID	No. core genes	No. accessory genes	No. unique genes	No. exclusively absent genes	N50	L50	ST <sup>b</sup>	No. genomes in NENT-DB
L28	2497	306	14	1	476,849	3	1	30
L44	2497	357	25	1	476,856	3	1	30
L42	2497	267	56	2	479,292	3	1	30
L128-3	2497	365	60	7	524,722	3	1	30
L174	2497	327	0	1	476,855	3	1	30
L49	2497	264	0	2	478,300	3	4	12
L58	2497	265	0	1	478,305	3	4	12
L105	2497	305	13	4	478,306	3	4	12
L124	2497	328	3	2	541,335	2	4	12
L12	2497	338	3	0	551,226	3	6	56
L164	2497	271	0	5	510,038	2	6	56
L52	2497	291	21	3	476,821	3	2332	0
L41	2497	294	37	3	478,264	3	59	4
L50	2497	246	32	0	609,560	2	7	9
L72	2497	379	10	2	446,908	3	7	9
L36	2497	299	13	3	571,154	2	451	1
L86	2497	352	111	4	582,512	2	91	1
L137	2497	295	0	0	1,490,250	1	29	1
L180	2497	331	43	1	1,489,724	1	29	1
L51	2497	321	0	0	1,497,622	1	37	8
L138	2497	354	12	1	1,530,524	1	37	8
L57	2497	275	0	2	543,302	2	412	4
L111	2497	277	0	0	543,302	2	412	4
L127	2497	276	0	0	526,717	2	412	4
L188	2497	277	0	0	543,302	2	412	4

**Table 3.** General features of the 25 *Listeria monocytogenes* draft genomes and number of comparable genomes available in the NENT database<sup>a</sup>. <sup>a</sup> database of the Swiss National Reference Centre for Enteropathogenic Bacteria and *Listeria* (NENT). <sup>b</sup>ST, sequence type; strains were assigned and grouped into STs in accordance with the BIGSdb-*L. monocytogenes* platform. For details see main text.



**Figure 2.** Minimum-spanning tree based on cgMLST allelic profiles of 25 *Listeria monocytogenes* isolated from surface waters. Each circle represents an allelic profile based on sequence analysis of 1,701 cgMLST target genes. The numbers on connecting lines represent the number of allelic differences between two strains. Each circle contains the strain ID, and CCs are color coded. Lineages and serotypes are indicated.



**Figure 3.** Minimum-spanning trees based on cgMLST allelic profiles of 25 *Listeria monocytogenes* isolated from surface water and 368 genome-sequenced clinical *L. monocytogenes* isolated during 2011–2020 available in the database of the Swiss National Reference Centre for Enteropathogenic Bacteria and *Listeria* (NENT) in Switzerland. Strains are grouped by clonal complex (CC). Each circle represents an allelic profile based on sequence analysis of 1,701 cgMLST target genes. The numbers on connecting lines represent the number of allelic differences between two strains. Clusters were defined as isolates containing  $\leq 10$  different alleles between a pair of isolates. Strains from this study are indicated with a pink star.

Internalin A, encoded by *inlA* is a key factor associated with pathogenic *L. monocytogenes*. Premature stop codons within *inlA*, often observed in food-associated and environmental strains, result in the presence of truncated InlA and reduced or loss of virulence<sup>30</sup>. In this study, all isolates contained full length *inlA* genes and were therefore potentially virulent. Among the 25 strains, there were several lineage and clonal complex specific SNP-introduced amino acid substitutions in *inlA* compared to the reference strain EDG-e. For example, the amino acid substitution R3K was only detected in strains belonging to lineage I, whilst Y774D was only detected in *L. monocytogenes* CC4 and CC6 (see Supplementary Table S2).

Further, *L. monocytogenes* CC6 (strains L12 and L164, respectively) all had a characteristic triple amino acid deletion in the pre-anchor region of InlA, the impact of which is not yet known<sup>30</sup> (see Supplementary Table S2).

In addition to *inlA*, the *L. monocytogenes* genomes in this study showed a variable combination of other internalin genes due to the presence or absence of *inlG* and *inlL* (see Supplementary Fig. S2). The *inlG* gene was lacking in lineage I strains, with the exception of two CC6 strains (L12 and L164). Similarly, *inlL* was absent in all lineage I strains as well as in three lineage II strains (L36, L86, and L180) (Supplementary Table 2 and Supplementary Fig. S2).

LIPI-1 is a PrfA dependent virulence gene cluster consisting of six genes (*prfA*, *plcA*, *hly*, *mpl*, *actA* and *plcB*) that are crucial for the infection cycle of *L. monocytogenes*<sup>31</sup>. In this study, LIPI-1 was present in all 25 strains (Supplementary Table 2 and Supplementary Fig. S2). All strains contained full length *prfA* genes (see Table S3). There were lineage and clonal complex specific SNP-introduced amino acid substitutions in *hly*, for example L35S and I438V were only detected in lineage I strains whilst V433I was unique to CC412 strains (see Supplementary Table S4).

There were various other minor lineage and strain specific genetic differences detected in further virulence factors including some amino acid changing SNPs in *plcA*, *plcB* and *actA*, but none that have previously been reported to alter functions of these proteins.

LIPI-3 encodes a biosynthetic cluster involved in the production of listeriolysin S (LLS), a hemolytic and cytotoxic factor postulated to play a role in *Listeria* gastrointestinal colonization<sup>32</sup>. LIPI-3 is strongly associated with lineage I strains and was present in all lineage I strains of this study with the exception of strain L41 which belonged to serotype 1/2b ST59 (Supplementary Table 2 and Supplementary Fig. S2).

LIPI-4 contains six genes encoding cellobiose-type phosphotransferase systems (PTS) that can enhance invasion, leading to neural and placental listeriosis<sup>6</sup>. The presence of LIPI-4 is strongly associated with hypervirulence of CC4 strains. Accordingly, LIPI-4 was identified among all CC4 strains in this study (Table 2 and Fig. S2).

SSI-1 is a five gene islet that contributes to the growth of *L. monocytogenes* under suboptimal conditions<sup>33,34</sup>. This islet has been previously shown to be a feature of *L. monocytogenes* CC7 and CC8 associated with persistence in food-processing plants but is also found in sporadic environmental strains<sup>26</sup>. In this study, SSI-1 was identified only in strains L50 and L72 both (CC7), and in L52 belonging to an ST2332 (Supplementary Table 2 and Supplementary Fig. S2).

Other VF including genes involved in adherence, intracellular survival, regulation of transcription and translation and surface protein anchoring were present in most of the strains, with some notable lineage, clonal complex, and serotype associated differences (see Supplementary Fig. S2). For instance, the adherence gene *ami*<sup>35</sup> was lacking in lineage I except in strain L41. Further, strains belonging to serotype 4b all contained *aut\_IVb* which is an allele of the invasion gene *aut*<sup>36</sup>, and all lacked *tagB*, a gene involved in teichoic acid biosynthesis<sup>37</sup>. All serotype 1/2a and 1/2b strains were as expected without the genes *gltA* and *gltB* which are serotype 4b-specific genes involved in teichoic acid biosynthesis<sup>38</sup>. Finally, all *L. monocytogenes* CC412 lacked the adherence gene *lapB*<sup>35</sup>.

**Identification of antimicrobial, heavy metal and disinfectant resistance determinants.** All 25 *L. monocytogenes* strains from this study contained four intrinsic antibiotic resistance genes, including the fosfomycin hydrolase gene *fosX*, the antibiotic efflux pump gene *lin*, the quinolone resistance efflux pump gene *norB*, and the sulfonamide resistance gene *sul*. Cadmium resistance genes *cadA1* and *cadC1* were detected in strain L86 (CC14). No arsenic or benzalkonium chloride resistance genes were detected (see Supplementary Fig. S2).

## Discussion

In this nationwide study, we recovered *L. monocytogenes* from water samples from rivers and streams localized within agricultural and nonagricultural environments, urbanized areas, and mountainous regions up to altitudes of 1560 m above sea level. The majority (21/84%) of the positive samples was retrieved within the geographical region of Switzerland that belongs to the so-called Central Plain. This area is characterized by settlement and urban areas, as well as agricultural areas, and represents the most densely populated region of Switzerland<sup>39</sup>. Further, with 76% of the positive samples located in the proximity to WWTP, anthropogenic sources of *L. monocytogenes* strains retrieved in this study appear likely, however, this observation needs to be confirmed by additional investigations that include further data collection. Indeed, previous studies suggest that *Listeria* species survive conventional wastewater treatment processes and that effluents of WWTPs are potential sources of clinically important *L. monocytogenes*<sup>40,41</sup>. However, the prevalence and diversities of the *L. monocytogenes* strains may have been influenced by the recovery procedure. In our study, the methodology included enrichment in HFB, as opposed to previous studies using other protocols to isolate *Listeria* from water samples, such as Universal Pre-enrichment Broth (UPB)<sup>21</sup>, or selective *Listeria* Enrichment Broth (LEB)<sup>15,17</sup>. It cannot be excluded that some lineages or serotypes of *L. monocytogenes* may differ in their ability to recover in HFB, and caution should be applied when comparing results obtained using different methodologies.

The prevalence of *L. monocytogenes* in surface water in this study was 13%. By comparison, *L. monocytogenes* was identified in 10% of surface water sampled in Canada<sup>15</sup>, in 13% in New York State<sup>42</sup>, 31% in Mid-Atlantic US<sup>21</sup>. Notably, *L. monocytogenes* was not detected in surface water in Austria<sup>13</sup>. However, prevalence of *L. monocytogenes* in natural water bodies may vary according to recovery methodology and sampling season<sup>20,21</sup>. Therefore, the lack of periodic sampling and the lack data for the summer and autumn seasons in this study could have influenced the results and it cannot be excluded that extending the study period to include warm seasons may have had an impact on prevalence and variety of the *L. monocytogenes* present in flowing surface waters. Therefore, our results may not apply directly to *L. monocytogenes* recovered from water during warm seasons. Nevertheless, the data from this study highlights the broad geographical distribution of clinically relevant *L. monocytogenes* in the aquatic ecosystem.

Among the isolates, the majority belonged to serotypes and clonal complexes corresponding to those from human listeriosis outbreaks and sporadic cases of human and animal infection. In our study, the majority of the strains were either serotype 1/2a (48%), or 4b (44%). These results are similar to those reported for surface waters analyzed in Canada, where serotypes 1/2a/3a and 4b/4d/4e constituted 49% and 32% of the isolates, respectively<sup>15</sup>. By contrast, other investigations found serotype 1/2a among 67% of water-derived *L. monocytogenes* in a further Canadian study, while *L. monocytogenes* recovered from water samples in California belonged predominantly to serotype 4b/4d/4e<sup>16</sup>. In spite of the limited number of isolates in the present study and the differences in methodologies and study settings used by previous investigators, these results of these earlier studies are supportive of our observation that *L. monocytogenes* populations in the aquatic environment contain serotypes that may cause listeriosis. Further, virulence gene profiling revealed that all the strains harbored intact genes associated with invasion and infection, underlining the virulence potential and clinical relevance of *L. monocytogenes* from the aquatic environment. These findings, although based on the analysis of a small number of isolates, are different from data on VFs found in many but not all isolates of serotypes 1/2a, 1/2b and 1/2c from food and food processing environments for example in Ireland and in the US, where *inlA* is truncated in 31% and in 45% of the isolates, respectively<sup>10,43</sup>. Our data are also supportive of a previous report regarding the integrity of the virulence gene *inlA* in *L. monocytogenes* recovered from natural waters<sup>17</sup>. Notably, there was a lack of benzalkonium and arsenic resistance genes and a very low prevalence of cadmium resistance genes among the strains in this study. These determinants are frequently associated with *L. monocytogenes* isolated from food and from humans, and their scarcity among the isolates in this study may reflect specific adaptations in the natural environment, consistent with a recent study that observed a very low prevalence of cadmium resistance genes among *L. monocytogenes* from wildlife<sup>44</sup>.

A considerable proportion (16%) of the clones belonged to CC4 which contains LIPI-4 and is considered hypervirulent based on its neurovirulence and capacity for placental infection<sup>6</sup>. CC4 is highly associated with



human isolates and, in contrast to other serotype 4b strains, to our knowledge has not been described in surface water so far<sup>45</sup>.

Likewise, CC1 and CC6 are major contributors to human listeriosis, and recent years have seen an increase of severe listeriosis cases related to *L. monocytogenes* CC6, a clone which was first implicated in a multistate outbreak in the US in 1998–1999<sup>46</sup>, and has since disseminated globally, causing one of the world's largest listeriosis outbreaks in South Africa in 2017–2018, a large outbreak in Germany during 2018–2019, a local outbreak in Switzerland in 2016<sup>47–49</sup>, and very recently, a nationwide outbreak in Switzerland in 2018–2020 that caused 34 cases and 10 deaths<sup>50</sup>.

The occurrence in surface water of *L. monocytogenes* belonging to CCs associated with disease highlights the potential of rivers, streams and inland canals as a reservoir for pathogenic *L. monocytogenes*. In particular, the use of river water for crop irrigation during dry seasons may allow *L. monocytogenes* in the water to enter the food chain. Irrigation has repeatedly been associated with an increased risk of pre-harvest produce contamination by *L. monocytogenes*, particularly if the irrigation water is drawn from surface water<sup>19,20,51,52</sup>. Further, irrigation within three days of harvest is associated with *L. monocytogenes* in produce production environments<sup>42</sup>. Therefore, river sourced irrigation water may indicate a public health risk, should contaminated product be consumed raw.

Similarly, the surface water samples in this study contained particular CCs that have reportedly caused bovine listeriosis, with *L. monocytogenes* CC1, CC4, and CC412 among the most common causes of ruminant encephalitis in central Europe<sup>28</sup>. These CCs are also frequently detected in the cattle farm environment including feed, drinking troughs, ruminant feces and manure<sup>25,28</sup>. However, the introduction routes of these CCs to the farm environment are currently not well understood, although spoiled silage is considered to be the primary source<sup>3,14</sup>. The occurrence of *L. monocytogenes* in rivers reported in this study points towards surface water a possible further source of clones causing disease in cattle. Notably, the use of river water for watering cattle may represent a possible exposure to bovine pathogenic *L. monocytogenes*.

## Conclusions

This study demonstrates that *L. monocytogenes* circulating in the aquatic environment belong to CCs and contain the same virulence traits as *L. monocytogenes* that are frequently isolated from human and animal clinical cases and from globally occurring outbreaks, including hypervirulent clones CC1, CC4, and CC6.

Our data contribute to a better understanding of the diversity of *L. monocytogenes* present in flowing surface waters. The results may provide information to improve crop irrigation strategies and cattle watering practices that aim to reduce the transmission of foodborne pathogens from surface water to fresh produce and to the farm environment.

## Material and methods

**Sampling.** A total of 191 water samples from different rivers and streams and inland canals throughout Switzerland were collected between January and May 2020. Sampling sites were located between 200 m and 1,730 m above sea level (see Supplementary Table S1). Water was taken from large rivers at 1 m depth using collection poles and sterile 500 mL containers. Smaller water bodies were sampled at 0.2–0.3 m depth using sterile 500 mL containers. The water samples were transported to the laboratory in a cool box. Each sample was stored at 4 °C for a maximum of 18 h. At each sampling site, weather conditions, ambient temperature and proximity to wastewater treatment plants were recorded.

**Bacterial isolation.** From each water sample 100 ml were passed through sterile, 0.22 µm membrane filters (Millipore, Billerica, MA, USA). For enrichment, the filters were incubated in 50 ml Half Frazer Broth (HFB, BioRad, Cressier, Switzerland) in sterile blender bags (Seward, Worthing, UK) at 30 °C for 48 h. One loopful each of the enriched cultures was streaked on Oxoid chromogenic Listeria agar (OCLA) plates (Oxoid, Pratteln, Switzerland) and incubated under aerobic conditions at 37 °C for 48 h. Presumptive *Listeria* colonies exhibiting green morphologies and opaque halos were subcultured on OCLA plates at 37 °C for 48 h. *L. monocytogenes* isolates were grown on sheep blood agar (Difco Laboratories) at 37 °C for 24 h, and kept at –80 °C in brain heart infusion (BHI) broth stocks (Oxoid, Hampshire, UK) containing 15% glycerol.

## Reference strains

To confirm lineages and serotypes, an average nucleotide identity (ANI) comparison was performed to create a phylogenetic tree that was calibrated with five *L. monocytogenes* genomes of known lineages and serotypes as reference strains (see Supplementary Fig. S1). The *L. monocytogenes* serotype 4b and 1/2b isolates from this study were compared to previously described outbreak strains LL195 (GenBank accession no. HF558398) and N2306 (CP011004). *L. monocytogenes* LL195 (CC1) was isolated from Swiss Vacherin Mont d'Or cheese during an outbreak 1983–1987<sup>53</sup>, and *L. monocytogenes* N2306 was detected in ready-to-eat salad during an outbreak 2013–2014<sup>54</sup>.

The *L. monocytogenes* serotype 1/2a isolates were compared to the reference strain *L. monocytogenes* EGD-e (NC003210), and to strains N1546 (CP013724) and N12-1273 (QYFZ00000000). *L. monocytogenes* EGD-e (CC9) was originally isolated from a rodent and is a widely used reference strain<sup>55</sup>. *L. monocytogenes* N1546 (CC8) is a clinical isolate recovered during the 2011 outbreak linked to ham products<sup>56,57</sup>. N12-1273 (CC412) is a previously characterized human listeriosis isolate<sup>49</sup>.

**Whole genome sequencing.** Genomic DNA was isolated using the DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany). Sequencing libraries were prepared using the Illumina Nextera DNA preparation kit and sequencing was performed on an Illumina MiSeq sequencer (Illumina, San Diego, CA, USA). Genomes

were sequenced with a minimal coverage of 50x. Following a quality assessment with FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>), the reads were assembled with Shovill 1.0.9 and Spades 3.12.0<sup>58</sup> and integrated into the Ridom SeqSphere+ software version 5.1.0 (Ridom, Münster, Germany)<sup>59</sup>.

**Serotyping and multilocus sequence typing.** In silico serotyping was performed in SeqSphere+ using gene targets described previously<sup>4</sup>. Classic MLST determination based on seven housekeeping genes were performed in accordance with the *L. monocytogenes* BIGSdb-*L. monocytogenes* platform (<https://bigsdbs.pasteur.fr/listeria>).

**Core genome MLST and Single-Nucleotide Polymorphism Analysis.** Core genome MLST (cgMLST) analysis was performed in accordance to the core genome defined by Ruppitsch et al.<sup>59</sup>. Sequences were blasted against 1701 genes of the reference genome of strain EGD-e, using the standard settings<sup>59</sup>. Cluster types (CTs) were determined upon submission to the *L. monocytogenes* cgMLST Ridom SeqSphere+ server (<http://www.cgmlst.org/ncs/schema/690488/>). Missing genes were ignored in all samples. Minimum spanning trees (MSTs) were generated in Ridom SeqSphere+ version 5.1.0 for visualization of strain relatedness. Clusters were defined as isolates containing  $\leq 10$  different alleles between a pair of isolates<sup>59</sup>.

Single-nucleotide polymorphism (SNP) analysis was performed using tparsnp in the harvest suite with option -c ignore MUMi activated<sup>60</sup>. An ANI was calculated based on MUMmer alignments as described previously<sup>61</sup>. Each isolate was compared to a reference strain of the same serotype, as listed above.

**Pan- and Core Genome Profiling.** The Bacterial Isolate Genome Sequence database (BIGSdb)<sup>62</sup> was used to generate the pan-genome of the 25 surface water isolates together with the five reference strains. The presence or absence of genes, including virulence genes, pathogenicity islands, antimicrobial resistance, heavy metal resistance, and biocide resistance genes across each genome was verified by manual curation using Basic Local Alignment Search Tool (BLAST) version 2.10.1 on a CLC genomics Workbench version 20.0.3.

**Geographical map.** Geospatial visualization was carried out by plotting GPS coordinates of the sampling sites onto a geographical map using the open source geographic information system (GIS) software QGIS (<https://qgis.org>).

## Data availability

This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under the accession numbers JACOE000000000-JACOFM000000000. The versions described in this paper are versions JACOE001000000-JACOFM010000000. Accession numbers for the individual *L. monocytogenes* strains from this study are listed in Table 2. The BioProject number is PRJNA657153.

Received: 16 November 2020; Accepted: 6 April 2021

Published online: 27 April 2021

## References

1. Matereke, L. T. & Okoh, A. I. *Listeria monocytogenes* virulence, antimicrobial resistance and environmental persistence: A review. *Pathogens* **9**, 528 (2020).
2. Allerberger, F. & Wagner, M. Listeriosis: A resurgent foodborne infection. *Clin. Microbiol. Infect.* **16**, 16–23 (2010).
3. Vázquez-Boland, J. A. et al. *Listeria* pathogenesis and molecular virulence determinants. *Clin. Microbiol. Rev.* **14**, 584–640 (2001).
4. Doumith, M. et al. Differentiation of the major *Listeria monocytogenes* serovars by multiplex PCR. *J. Clin. Microbiol.* **42**, 3819–3822 (2004).
5. Orsi, R. H. et al. *M. Listeria monocytogenes* lineages: Genomics, evolution, ecology, and phenotypic characteristics. *Int. J. Med. Microbiol.* **301**, 79–96 (2011).
6. Maury, M. M. et al. Uncovering *Listeria monocytogenes* hypervirulence by harnessing its biodiversity. *Nat. Genet.* **48**, 308–313 (2016).
7. Gözel, B. et al. Hyperinvasiveness of *Listeria monocytogenes* sequence type 1 is independent of lineage I-specific genes encoding internalin-like proteins. *MicrobiologyOpen* **8**, e790 (2019).
8. Travier, L. & Lecuit, M. *Listeria monocytogenes* ActA: A new function for a 'classic' virulence factor. *Curr. Opin. Microbiol.* **17**, 53–60 (2014).
9. Johansson, J. & Freitag, N. E. Regulation of *Listeria monocytogenes* virulence. In *Gram-Positive Pathogens* (ed. Richard, P.) 836–850 (ASM Press, Washington, 2019).
10. Hurlley, D. et al. Whole-genome sequencing-based characterization of 100 *Listeria monocytogenes* isolates collected from food processing environments over a four-year period. *mSphere* **4**, e00252-19 (2019).
11. Nightingale, K. K. et al. *inlA* premature stop codons are common among *Listeria monocytogenes* isolates from foods and yield virulence-attenuated strains that confer protection against fully virulent strains. *Appl. Environ. Microbiol.* **74**, 6570–6583 (2008).
12. Nightingale, K. K. et al. Ecology and transmission of *Listeria monocytogenes* infecting ruminants and in the farm environment. *Appl. Environ. Microbiol.* **70**, 4458–4467 (2004).
13. Linke, K. et al. Reservoirs of *Listeria* species in three environmental ecosystems. *Appl. Environ. Microbiol.* **80**, 5583–5592 (2014).
14. Oevermann, A. et al. Rhombencephalitis caused by *Listeria monocytogenes* in humans and ruminants: A zoonosis on the rise. *Interdiscip. Perspect. Infect. Dis.* **2010**, 632513 (2010).
15. Lytautey, E. et al. Distribution and characteristics of *Listeria monocytogenes* isolates from surface waters of the South Nation River watershed, Ontario, Canada. *Appl. Environ. Microbiol.* **73**, 5401–5410 (2007).
16. Stea, E. C., Purdue, L. M., Jamieson, R. C., Yost, C. K. & Truelstrup Hansen, L. Comparison of the prevalences and diversities of *Listeria* species and *Listeria monocytogenes* in an urban and a rural agricultural watershed. *Appl. Environ. Microbiol.* **81**(11), 3812–3822 (2015).
17. Gorski, L. et al. The majority of genotypes of the virulence gene *inlA* are intact among natural watershed isolates of *Listeria monocytogenes* from the central California coast. *PLoS ONE* **11**, e0167566 (2016).

18. Sauders, B. D. *et al.* Diversity of *Listeria* species in urban and natural environments. *Appl. Environ. Microbiol.* **78**, 4420–4433 (2012).
19. Strawn, L. K. *et al.* Risk factors associated with *Salmonella* and *Listeria monocytogenes* contamination of produce fields. *Appl. Environ. Microbiol.* **79**, 7618–7627 (2013).
20. Weller, D. *et al.* Complex interactions between weather, and microbial and physicochemical water quality impact the likelihood of detecting foodborne pathogens in agricultural water. *Front. Microbiol.* **11**, 134 (2020).
21. Sharma, M. *et al.* Prevalence of *Salmonella* and *Listeria monocytogenes* in non-traditional irrigation waters in the Mid-Atlantic United States is affected by water type, season, and recovery method. *PLoS ONE* **15**, e0229365 (2020).
22. Papić, B. *et al.* Source tracking on a dairy farm reveals a high occurrence of subclinical mastitis due to hypervirulent *Listeria monocytogenes* clonal complexes. *J. Appl. Microbiol.* **127**, 1349–1361 (2019).
23. Chen, Y. *et al.* Core genome multilocus sequence typing for identification of globally distributed clonal groups and differentiation of outbreak strains of *Listeria monocytogenes*. *Appl. Environ. Microbiol.* **82**, 6258–6272 (2016).
24. Koopmans, M. M. *et al.* *Listeria monocytogenes* sequence type 6 and increased rate of unfavorable outcome in meningitis: Epidemiologic cohort study. *Clin. Infect. Dis.* **57**, 247–253 (2013).
25. Papić, B. *et al.* Genetic diversity of *Listeria monocytogenes* strains in ruminant abortion and rhombencephalitis cases in comparison with the natural environment. *BMC Microbiol.* **19**, 299 (2019).
26. Knudsen, G. M. *et al.* Genome-wide analyses of *Listeria monocytogenes* from food-processing plants reveal clonal diversity and date the emergence of persisting sequence types. *Environ. Microbiol. Rep.* **9**, 428–440 (2017).
27. den Bakker, H. C. *et al.* Multilocus sequence typing of outbreak-associated *Listeria monocytogenes* isolates to identify epidemic clones. *Foodborne Pathog. Dis.* **7**, 257–265 (2010).
28. Dreyer, M. *et al.* *Listeria monocytogenes* sequence type 1 is predominant in ruminant rhombencephalitis. *Sci. Rep.* **6**, 36419 (2016).
29. Kim, S. W. *et al.* Genetic diversity and virulence profiles of *Listeria monocytogenes* recovered from bulk tank milk, milk filters, and milking equipment from dairies in the United States (2002 to 2014). *PLoS ONE* **13**, e0197053 (2018).
30. Cantinelli, T. *et al.* “Epidemic clones” of *Listeria monocytogenes* are widespread and ancient clonal groups. *J. Clin. Microbiol.* **51**, 3770–3779 (2013).
31. Moura, A. *et al.* Whole genome-based population biology and epidemiological surveillance of *Listeria monocytogenes*. *Nat. Microbiol.* **2**, 16185 (2016).
32. Cotter, P. D. *et al.* Listeriolysin S, a novel peptide haemolysin associated with a subset of lineage I *Listeria monocytogenes*. *PLoS Pathog.* **4**, e1000144 (2008).
33. Ryan, S. *et al.* A five-gene stress survival islet (SSI-1) that contributes to the growth of *Listeria monocytogenes* in suboptimal conditions. *J. Appl. Microbiol.* **109**, 984–995 (2010).
34. Malekmohammadi, S. *et al.* Genetic and environmental factors influence *Listeria monocytogenes* nisin resistance. *J. Appl. Microbiol.* **123**, 262–270 (2017).
35. Camejo, A. *et al.* The arsenal of virulence factors deployed by *Listeria monocytogenes* to promote its cell infection cycle. *Virulence* **2**, 379–394 (2011).
36. Cabanes, D., Dussurget, O., Dehoux, P. & Cossart, P. Auto, a surface associated autolysin of *Listeria monocytogenes* required for entry into eukaryotic cells and virulence. *Mol. Microbiol.* **51**, 1601–1614 (2004).
37. Formstone, A., Carballido-López, R., Noirot, P., Errington, J. & Scheffers, D. J. Localization and interactions of teichoic acid synthetic enzymes in *Bacillus subtilis*. *J. Bacteriol.* **190**(5), 1812–1821 (2008).
38. Lei, X. H., Fiedler, F., Lan, Z. & Kathariou, S. A novel serotype-specific gene cassette (*glta-gltb*) is required for expression of teichoic acid-associated surface antigens in *Listeria monocytogenes* of serotype 4b. *J. Bacteriol.* **183**, 1133–1139 (2001).
39. Swiss Federal Statistics Office. *Land use in Switzerland: Results of the Swiss land use statistics*. (ed. SFSO) (Neuchâtel, 2013).
40. Odadjare, E. E. O. & Okoh, A. I. Prevalence and distribution of *Listeria* pathogens in the final effluents of a rural wastewater treatment facility in the eastern cape province of South Africa. *World J. Microbiol. Biotechnol.* **26**, 297–307 (2010).
41. Paillard, D. *et al.* Occurrence of *Listeria* spp. in effluents of French urban wastewater treatment plants. *Appl. Environ. Microbiol.* **71**, 7562–7566 (2005).
42. Weller, D., Wiedmann, M. & Strawn, L. K. Irrigation is significantly associated with an increased prevalence of *Listeria monocytogenes* in produce production environments in New York State. *J. Food Prot.* **78**, 1132–1141 (2015).
43. Kovacevic, J. *et al.* Examination of food chain-derived *Listeria monocytogenes* strains of different serotypes reveals considerable diversity in *inlA* genotypes, mutability, and adaptation to cold temperatures. *Appl. Environ. Microbiol.* **79**, 1915–1922 (2013).
44. Parsons, C. *et al.* *Listeria monocytogenes* at the human-wildlife interface: black bears (*Ursus americanus*) as potential vehicles for *Listeria*. *Microb. Biotechnol.* **13**, 706–721 (2020).
45. Lee, S. *et al.* *Listeria monocytogenes* source distribution analysis indicates regional heterogeneity and ecological niche preference among serotype 4b clones. *MBio* **9**, 5. <https://doi.org/10.1128/mBio.00396-18> (2018).
46. Centers for Disease Control and Prevention. Update: Multistate outbreak of listeriosis—United States, 1998–1999. *Morb. Mortal. Wkly. Rep.* **47**, 1117–1118 (1999).
47. Smith, A. M. *et al.* Outbreak of *Listeria monocytogenes* in South Africa, 2017–2018: Laboratory activities and experiences associated with whole-genome sequencing analysis of isolates. *Foodborne Pathog. Dis.* **16**, 524–530 (2019).
48. Halbedel, S. *et al.* Large nationwide outbreak of invasive listeriosis associated with blood sausage, Germany, 2018–2019. *Emerg. Infect. Dis.* **26**, 1456–1464 (2020).
49. Althaus, D. *et al.* Characterization of *Listeria monocytogenes* strains isolated during 2011–2013 from human infections in Switzerland. *Foodborne Pathog. Dis.* **11**, 753–758 (2014).
50. Nüesch-Inderbinen, M. *et al.* Listeriosis caused by persistence of *Listeria monocytogenes* serotype 4b sequence type 6 in cheese production environment. *Emerg. Infect. Dis.* **27**, 284–288 (2021).
51. Weller, D., Wiedmann, M. & Strawn, L. K. Spatial and temporal factors associated with an increased prevalence of *Listeria monocytogenes* in spinach fields in New York State. *Appl. Environ. Microbiol.* **81**, 6059–6069 (2015).
52. Jongman, M. & Korsten, L. Assessment of irrigation water quality and microbiological safety of leafy greens in different production systems. *J. Food Saf.* **37**, e12324 (2017).
53. Weinmaier, T. *et al.* Complete genome sequence of *Listeria monocytogenes* LL195, a serotype 4b strain from the 1983–1987 listeriosis epidemic in Switzerland. *Genome Announc.* **1**, e00152-12 (2013).
54. Stephan, R. *et al.* Foodborne transmission of *Listeria monocytogenes* via ready-to-eat salad: a nationwide outbreak in Switzerland, 2013–2014. *Food Control* **57**, 14–17 (2015).
55. Glaser, P. *et al.* Comparative genomics of *Listeria* species. *Science* **294**, 849–852 (2001).
56. Tasara, T., Klumpp, J., Bille, J. & Stephan, R. Genome sequences of *Listeria monocytogenes* strains responsible for cheese- and cooked ham product-associated swiss listeriosis outbreaks in 2005 and 2011. *Genome Announc.* **4**, e00106-e116 (2016).
57. Hächler, H. *et al.* Outbreak of listeriosis due to imported cooked ham, Switzerland 2011. *Euro. Surveill.* **18**, 20469 (2013).
58. Bankevich, A. *et al.* SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol.* **19**, 455–477 (2012).
59. Ruppitsch, W. *et al.* Defining and evaluating a core genome multilocus sequence typing scheme for whole-genome sequence-based typing of *Listeria monocytogenes*. *J. Clin. Microbiol.* **53**, 2869–2876 (2015).
60. Treangen, T. J., Ondov, B. D., Koren, S. & Phillippy, A. M. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. *Genome Biol.* **15**, 524 (2014).



61. Stevens, M. J. A. *et al.* Whole-genome-based phylogeny of *Bacillus cytotoxicus* reveals different clades within the species and provides clues on ecology and evolution. *Sci. Rep.* **9**, 1984 (2019).
62. Jolley, K. A. & Maiden, M. C. BIGSdb: Scalable analysis of bacterial genome variation at the population level. *BMC Bioinform.* **11**, 595 (2010).

## Acknowledgements

This work was supported in part by the Swiss Federal Office of Public Health, Division Communicable Diseases.

## Author contributions

The study was designed by R.S. Sampling was accomplished by S.R. Geodata processing was carried out by K.Z. Microbiological analyses were performed by S.R and K.Z. WGS was done by N.C. Bioinformatic analyses were conducted by M.J.A.S and F.M. Data analyses was conducted by S.R., F.M., M.N.I., and R.S. The manuscript was written by M.N.-I. S.R., R.S., and M.J.A.S. contributed to writing and revising the manuscript. All authors agreed on the final version.

## Competing interests

The authors declare no competing interests.

## Additional information

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1038/s41598-021-88514-y>.

**Correspondence** and requests for materials should be addressed to M.N.-I.

**Reprints and permissions information** is available at [www.nature.com/reprints](http://www.nature.com/reprints).

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021

## **4 Anhänge**

Supplementary information

## Supplementary Information

### **Environmental dissemination of pathogenic *Listeria monocytogenes* in flowing surface waters in Switzerland**

Susanne Raschle,<sup>a</sup> Roger Stephan,<sup>a</sup> Marc J.A. Stevens,<sup>a</sup> Nicole Cernela,<sup>a</sup> Katrin Zurfluh,<sup>a</sup> Francis Muchaamba,<sup>a</sup> Magdalena Nüesch-Inderbinen<sup>a</sup> \*

<sup>a</sup>Institute for Food Safety and Hygiene, Vetsuisse Faculty, University of Zurich, Zurich, Switzerland

\* Corresponding author: [magdalena.nueesch-inderbinen@uzh](mailto:magdalena.nueesch-inderbinen@uzh).

**Table S1:** Geographical features of water samples collected from surface water bodies throughout Switzerland, 2020.

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L_1	14.01.20	47°36'23" 9°6'40"	fair (0-5°C)	470	stream	Chemibach	+
L_2	14.01.20	47°35'39" 8°57'55"	fair (0-5°C)	400	stream	Chemebach	+
L_3	14.01.20	47°32'29" 8°54'27"	fair (0-5°C)	430	river	Murg	+
L_4	14.01.20	47°29'33" 8°59'0"	fair (0-5°C)	510	river	Murg	+
L_5	14.01.20	47°34'6" 9°6'23"	fair (0-5°C)	430	river	Giessen	+
L_6	14.01.20	47°38'19" 9°13'8"	fair (0-5°C)	410	stream	Töbelibach	-
L_7	14.01.20	47°35'18" 8°56'27"	fair (0-5°C)	390	river	Thur	-
L_8	16.01.20	47°26'27" 8°42'13"	fair (-2-8°C)	480	river	Kempt	+
L_9	16.01.20	47°22'15" 8°48'34"	fair (-2-8°C)	620	river	Luppen	+
L_10	16.01.20	47°15'13" 8°47'46"	fair (-2-8°C)	490	stream	Klausbach	+
L_11	16.01.20	47°17'45" 8°43'13"	fair (-2-8°C)	450	stream	Lieburgerbach	+
L_12	16.01.20	47°19'42" 8°36'54"	fair (-2-8°C)	580	stream	Chliweidlibach	+
L_13	16.01.20	47°31'6" 8°39'17"	fair (-2-8°C)	410	river	Töss	+
L_14	20.01.20	47°39'42" 8°58'17"	fair (-2-1°C)	400	stream	Feldbach	+
L_15	20.01.20	47°38'20" 8°46'25"	fair (-2-1°C)	430	stream	Mülibach	+
L_16	20.01.20	47°45'14" 8°41'37"	fair (-2-1°C)	440	river	Biber	-
L_17	20.01.20	47°41'18" 8°27'14"	fair (-2-1°C)	410	inland canal	Klingengraben	+
L_18	20.01.20	47°34'10" 8°28'47"	fair (-2-1°C)	350	river	Glatt	+
L_19	20.01.20	47°35'57" 8°17'43"	fair (-2-1°C)	320	river	Rhein	+
L_20	20.01.20	47°29'6" 8°17'49"	fair (-2-1°C)	350	river	Limmat	-
L_21	20.01.20	47°29'2" 8°12'52"	fair (-2-1°C)	340	river	Aare	+
L_22	20.01.20	47°27'33" 8°14'42"	fair (-2-1°C)	340	river	Reuss	+
L_23	20.01.20	47°31'7" 8°0'55"	fair (-2-1°C)	340	stream	Sissle	+
L_24	20.01.20	47°24'46" 8°5'4"	fair (-2-1°C)	370	river	Aare	+
L_25	27.01.20	47°34'36" 7°50'26"	cloudy (1-3°C)	290	stream	Möhlin	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L_26	27.01.20	47°33'11" 7°47'41"	cloudy (1-3°C)	280	stream	Magdenbach	-
L_27	27.01.20	47°33'5" 7°45'55"	cloudy (1-3°C)	270	river	Rhein	+
L_28	27.01.20	47°30'56" 7°43'14"	cloudy (1-3°C)	290	river	Ergolz	+
L_29	27.01.20	47°35'18" 7°35'22"	cloudy (1-3°C)	250	river	Rhein	+
L_30	27.01.20	47°32'47" 7°37'23"	cloudy (1-3°C)	260	stream	Birs	+
L_31	27.01.20	47°25'41" 7°26'42"	cloudy (1-3°C)	420	stream	Lützel	+
L_32	27.01.20	47°21'54" 7°21'1"	cloudy (1-3°C)	410	stream	La Sorne	-
L_33	27.01.20	47°21'31" 7°8'19"	cloudy (1-3°C)	440	stream	Risseau du Doubs	+
L_34	27.01.20	47°25'59" 7°4'49"	cloudy (1-3°C)	410	stream	Lällaine	+
L_35	27.01.20	47°16'42" 7°23'48"	cloudy (1-3°C)	550	stream	La Raus	-
L_36	27.01.20	47°16'49" 7°31'41"	cloudy (1-3°C)	680	stream	Dünner	+
L_37	27.01.20	47°77'26" 8°27'74"	cloudy (1-3°C)	1893	stream	Tannenbach	-
L_38	09.01.20	46°36'47" 9°35'21"	fair (-8-2°C)	1330	river	Julia	+
L_39	09.01.20	46°44'34" 9°25'55"	fair (-8-2°C)	630	river	Hinterrhein	+
L_40	09.01.20	46°49'29" 9°24'26"	fair (-8-2°C)	580	river	Vorderrhein	-
L_41	11.02.20	47°33'6" 9°19'34"	heavy rain (-1-5°C)	420	stream	Aach	+
L_42	11.02.20	47°29'56" 9°13'51"	heavy rain (-1-5°C)	470	river	Thur	+
L_43	11.02.20	47°24'52" 9°12'3"	heavy rain (-1-5°C)	570	river	Glatt	+
L_44	11.02.20	47°19'25" 9°5'6"	heavy rain (-1-5°C)	600	stream	Lederbach	-
L_45	11.02.20	47°13'30" 9°11'20"	heavy rain (-1-5°C)	730	river	Thur	+
L_46	11.02.20	47°19'44" 9°17'43"	heavy rain (-1-5°C)	800	stream	Urnäsch	+
L_47	11.02.20	47°20'49" 9°23'48"	heavy rain (-1-5°C)	780	river	Sitter	+
L_48	11.02.20	47°22'12" 9°34'11"	heavy rain (-1-5°C)	420	stream	Rietaach	+
L_49	11.02.20	47°29'31" 9°33'56"	heavy rain (-1-5°C)	400	river	Rhein	+
L_50	11.02.20	47°21'16" 9°7'45"	heavy rain (-1-5°C)	630	river	Necker	+
L_51	11.02.20	47°22'0" 9°24'9"	heavy rain (-1-5°C)	800	stream	Rotbach	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L_52	11.02.20	47°33'23" 9°22'42"	heavy rain (-1-5°C)	400	river	Aach	+
L_53	17.02.20	47°19'35" 7°50'32"	cloudy (6-12°C)	420	river	Dünner	+
L_54	17.02.20	47°18'54" 7°53'43"	cloudy (6-12°C)	400	river	Wigger	-
L_55	17.02.20	47°15'3" 7°46'14"	cloudy (6-12°C)	430	river	Aare	+
L_56	17.02.20	47°12'17" 7°41'52"	cloudy (6-12°C)	450	stream	Önz	+
L_57	17.02.20	47°6'35" 7°32'8"	cloudy (6-12°C)	480	stream	Aefligen	+
L_58	17.02.20	47°1'28" 7°38'52"	cloudy (6-12°C)	560	river	Emme	+
L_59	17.02.20	46°56'55" 7°45'23"	cloudy (6-12°C)	650	stream	Ilfis	+
L_60	17.02.20	46°54'18" 7°55'51"	cloudy (6-12°C)	830	stream	Lombach	-
L_61	17.02.20	47°1'25" 8°4'2"	cloudy (6-12°C)	610	river	Kleine Emme	+
L_62	17.02.20	47°5'55" 7°57'31"	cloudy (6-12°C)	620	stream	Nollentalbach	-
L_63	17.02.20	47°13'57" 7°58'12"	rainy (6-12°C)	460	stream	Wigger	+
L_64	16.02.20	47°6'9" 9°20'6"	fair (6-12°C)	435	stream	Schils	+
L_65	16.02.20	47°6'15" 9°20'5"	fair (6-12°C)	430	stream	Seez	+
L_66	16.02.20	47°5'52" 9°18'21"	fair (6-12°C)	960	stream	Tobelbach	-
L_67	16.02.20	47°4'59" 9°19'3"	fair (6-12°C)	950	stream	Ruslenbach	-
L_68	16.02.20	46°54'59" 9°45'29"	fair (6-12°C)	790	river	Landquart	+
L_69	17.02.20	47°6'1" 7°57'45"	cloudy (6-12°C)	610	stream	Wigger	-
L_70	24.02.20	47°18'58" 8°3'5"	cloudy (12-15° C)	450	river	Suhre	+
L_71	24.02.20	47°13'41" 8°4'32"	cloudy (12-15° C)	480	river	Suhre	+
L_72	24.02.20	47°17'21" 8°8'1"	cloudy (12-15° C)	490	stream	Wyna	+
L_73	24.02.20	47°10'43" 8°16'39"	cloudy (12-15° C)	470	inland canal	Ron	+
L_74	24.02.20	47°4'15" 8°17'32"	cloudy (12-15° C)	430	river	Reuss	+
L_75	24.02.20	46°56'46" 8°16'0"	cloudy (12-15° C)	430	river	Sarner Aa	+
L_76	24.02.20	46°58'17" 8°20'26"	fair (12-18°C)	440	inland canal	Vorlauter	+
L_77	24.02.20	46°58'39" 8°25'3"	fair (12-18°C)	440	stream	Engelburger Aa	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L_78	24.02.20	46°57'30" 8°31'33"	fair (12-18°C)	720	stream	Choltalbach	-
L_79	24.02.20	46°53'17" 8°37'6"	fair (12-18°C)	440	inland canal	Giessenkanal	+
L_80	24.02.20	46°53'0" 8°37'7"	fair (12-18°C)	440	river	Reuss	-
L_81	24.02.20	47°0'45" 8°37'10"	fair (12-18°C)	440	stream	Seeweren	+
L_82	24.02.20	47°6'41" 8°27'4"	fair (12-18°C)	430	stream	Elibach	-
L_83	24.02.20	47°11'58" 8°26'27"	fair (12-18°C)	430	stream	Bach Lorze	-
L_84	24.02.20	47°12'9" 8°25'57"	fair (12-18°C)	390	inland canal	Kanal Lorze	+
L_85	24.02.20	47°0'42" 8°37'55"	fair (12-18°C)	440	river	Muota	-
L_86	02.03.20	47°13'14" 7°34'25"	cloudy (2-8°C)	420	river	Aare	+
L_87	02.03.20	47°10'21" 7°25'14"	cloudy (2-8°C)	430	river	Aare	+
L_88	02.03.20	47°10'33" 7°24'59"	cloudy (2-8°C)	430	stream	Witibachkanal	+
L_89	02.03.20	47°7'10" 7°15'31"	cloudy (2-8°C)	410	inland canal	Büren	+
L_90	02.03.20	47°11'16" 7°12'17"	cloudy (2-8°C)	630	stream	Schüss	+
L_91	02.03.20	47°13'32" 7°7'48"	cloudy (2-8°C)	860	stream	Trame	+
L_92	02.03.20	47°9'47" 7°1'56"	cloudy (2-8°C)	730	stream	Schüss	+
L_93	02.03.20	47°7'35" 6°51'25"	cloudy (2-8°C)	960	stream	la Ronde	+
L_94	02.03.20	47°1'10" 6°53'46"	cloudy (2-8°C)	730	stream	la Sagnetanna	+
L_95	02.03.20	47°0'38" 7°1'55"	cloudy (2-8°C)	430	inland canal	Zihlkana	+
L_96	02.03.20	46°58'0" 7°10'52"	cloudy (2-8°C)	430	inland canal	Maria Brunnenbach	+
L_97	02.03.20	47°5'32" 7°18'28"	cloudy (2-8°C)	440	river	Alte Aare	+
L_98	02.03.20	46°57'1" 7°9'43"	cloudy (2-8°C)	440	inland canal	Galmizkanal	+
L_99	02.03.20	46°42'16" 8°51'41"	fair (0-7°C)	1100	stream	Acletta	+
L_100	02.03.20	46°46'49" 9°13'49"	fair (0-7°C)	680	river	Vorderrhein	+
L_101	02.03.20	47°10'35" 8°54'39"	rainy (3-7°C)	440	stream	Färlibach	-
L102	09.03.20	47°26'17" 8°33'29"	rain (4-7°C)	420	river	Glatt	+
L103	09.03.20	47°27'2" 8°25'12"	rain (4-7°C)	420	stream	Furtbach	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L104	09.03.20	47°24'19" 8°28'13"	rain (4-7°C)	390	river	Limmat	+
L105	09.03.20	47°21'21" 8°20'45"	rain (4-7°C)	370	river	Reuss	+
L106	09.03.20	47°21'49" 8°25'9"	rain (4-7°C)	450	stream	Reppisch	+
L107	09.03.20	47°17'28" 8°25'59"	rain (4-7°C)	460	stream	Jonen	+
L108	09.03.20	47°12'51" 8°34'53"	rain (4-7°C)	540	stream	Sarbach	+
L109	09.03.20	47° 8'34" 8°45'5"	rain (4-7°C)	860	river	Alp	+
L110	09.03.20	47°10'54" 8°57'42"	rain (4-7°C)	410	inland canal	Wildbachkanal	+
L111	09.03.20	47°10'3" 9°0'51"	rain (4-7°C)	430	inland canal	Linthkanal	+
L112	09.03.20	47°5'53" 9°3'49"	rain (4-7°C)	430	stream	Mülibach	-
L113	09.03.20	47°3'31" 9°3'8"	rain (4-7°C)	460	stream	Löntsch	-
L114	09.03.20	47°2'35" 9°4'17"	rain (4-7°C)	470	river	Linth	-
L115	09.03.20	46°59'3" 9°8'41"	rain (4-7°C)	770	river	Senft	+
L116	09.03.20	46°56'14" 9°0'55"	rain (4-7°C)	590	river	Linth	-
L117	09.03.20	47°14'2" 8°55'29"	rain (4-7°C)	400	stream	Wagnerbach	+
L118	08.03.20	46°42'41" 9°32'22"	rain (4-7°C)	1600	stream	Sporz	-
L119	08.03.20	46°42'31" 9°33'7"	rain (4-7°C)	1400	stream	Rain digl Lai	+
L120	08.03.20	46°45'14" 9°47'12"	rain (4-7°C)	1440	river	Landwasser	+
L121	09.03.20	47°25'33" 8°32'30"	rain (4-7°C)	430	stream	Chatzenbach	-
L122	15.03.20	46°30'49" 9°51'47"	fair (2-13°C)	1710	river	Inn	+
L123	15.03.20	46°30'30" 9°52'53"	fair (2-13°C)	1730	river	Flaz repp	-
L124	15.03.20	46°57'43" 10°24'53"	fair (2-13°C)	1560	stream	Schergenbach	-
L125	15.03.20	46°47'57" 10°19'9"	fair (2-13°C)	1170	river	Inn	+
L126	15.03.20	46°46'1" 10°6'37"	fair (2-13°C)	1420	stream	Lavinuoz	-
L127	16.03.20	46°57'14" 6°43'35"	fair (-1-10°C)	730	stream	Noirigue	+
L128	16.03.20	46°54'60" 6°36'39"	fair (-1-10°C)	730	river	Areuse	+
L129	16.03.20	46°48'32" 6°33'1"	fair (-1-10°C)	580	stream	L'Arnon	+



Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L130	16.03.20	46°47'3" 6°35'6"	fair (-1-10°C)	430	stream	le Bey	+
L131	16.03.20	46°42'56" 6°23'21"	fair (-1-10°C)	740	river	l'Orbre	+
L132	16.03.20	46°36'11" 6°14'25"	fair (-1-10°C)	1010	river	l'Orbre	+
L133	16.03.20	46°24'49" 6°15'20"	fair (-1-10°C)	430	river	Promenthoux	-
L134	16.03.20	46°16'35" 6°9'59"	fair (-1-10°C)	380	stream	la Versoix	-
L135	16.03.20	46°11'40" 6°5'19"	fair (-1-10°C)	350	river	Rhone	+
L136	16.03.20	46°10'36" 6°0'28"	fair (-1-10°C)	350	river	Rhone	+
L137	16.03.20	46°10'57" 6°0'36"	fair (-1-16°C)	350	river	Allondon	-
L138	16.03.20	46°10'59" 6°11'1"	fair (-1-16°C)	400	stream	Seymaz	-
L139	16.03.20	46°10'45" 6°10'54"	fair (-1-16°C)	390	river	Arve	+
L140	16.03.20	46°32'36" 6°33'2"	fair (-1-16°C)	390	river	Venoge	+
L141	16.03.20	46°38'26" 6°37'29"	fair (-1-16°C)	600	river	le Talent	+
L142	16.03.20	46°50'7" 6°56'16"	fair (-1-16°C)	450	stream	Broye	+
L143	16.03.20	46°47'3" 7°6'57"	fair (-1-16°C)	580	river	la Glane	+
L144	17.03.20	46°48'54" 7°9'52"	fair (-1-8°C)	535	river	Saane	+
L145	17.03.20	46°55'5" 7°14'23"	fair (-1-8°C)	480	river	Saane	+
L146	17.03.20	46°54'15" 7°14'9"	fair (-1-8°C)	490	river	Sense	-
L147	17.03.20	46°58'23" 7°25'40"	fair (-1-8°C)	470	river	Aare	+
L148	17.03.20	46°52'48" 7°32'45"	fair (-1-8°C)	520	river	Giessen	+
L149	17.03.20	46°46'52" 7°35'56"	fair (5-18°C)	550	river	Aare	+
L150	17.03.20	46°39'14" 7°34'32"	fair (5-18°C)	670	stream	Simmen	-
L151	17.03.20	46°35'51" 7°39'39"	fair (5-18°C)	750	river	Entschligen	+
L152	17.03.20	46°40'59" 7°39'35"	fair (5-18°C)	620	river	Kander	-
L153	17.03.20	46°39'11" 7°52'12"	fair (5-18°C)	610	river	Lütschine	-
L154	17.03.20	46°40'28" 7°50'36"	fair (5-18°C)	560	river	Aare	+
L155	17.03.20	46°43'59" 8°9'37"	fair (5-18°C)	600	river	Aare	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L156	17.03.20	46°49'57" 8° 10'45"	fair (5-18°C)	480	stream	Lau	-
L157	03.05.20	47°13'2" 9°29'60"	fair (4-14°C)	440	river	Rhein	+
L158	03.05.20	47°1'24" 9°30'0"	fair (4-14°C)	500	river	Rhein	+
L159	03.05.20	46°52'18" 9°31'42"	fair (4-14°C)	510	river	Rhein	+
L160	03.05.20	46°33'18" 9°19'55"	fair (1-12°C)	1440	river	Hinterrhein	+
L161	03.05.20	46°17'35" 9°10'48"	fair (6-20°C)	400	river	Moesa	+
L162	03.05.20	46°10'30" 8°59'28"	fair (6-20°C)	220	river	Ticino	+
L163	03.05.20	46°2'8" 8°58'20"	fair (6-20°C)	320	river	Cassarate	+
L164	03.05.20	46°0'23" 8°54'48"	fair (6-20°C)	280	river	Veduggio	+
L165	03.05.20	45°53'33" 8°58'24"	fair (6-20°C)	280	river	Laveggio	+
L166	03.05.20	45°50'31" 9°2'13"	fair (6-24°C)	230	river	Breggia	+
L167	03.05.20	46°10'15" 8°51'31"	fair (6-24°C)	200	river	Versasca	-
L168	03.05.20	46°10'47" 8° 45'3"	fair (6-24°C)	220	river	Melezza	-
L169	03.05.20	46°16'48" 8°39'59"	fair (6-24°C)	360	river	Maggia	-
L170	03.05.20	46°19'52" 8°58'29"	fair (6-24°C)	270	river	Ticino	+
L171	03.05.20	46°29'21" 8°44'39"	fair (6-20°C)	940	river	Ticino	+
L172	03.05.20	46°31'29" 8°37'36"	Fair(6-18°C)	1100	river	Canaria	+
L173	03.05.20	46°46'19" 8°40'11"	fair (5-15°C)	510	river	Kärstelenbach	-
L174	12.05.20	46°38'14" 7°3'20"	light rain (3-6°C)	730	stream	La Sionge	-
L175	12.05.20	46°36'40" 7°5'29"	light rain (3-6°C)	680	river	Saane	+
L176	12.05.20	46°28'8" 6°50'53"	light rain (3-6°C)	400	river	la Veveyse	-
L177	12.05.20	46°22'30" 6°55'45"	light rain (3-6°C)	380	stream	Eau Froide	+
L178	12.05.20	46°13'2 7°0'29"	light rain (3-6°C)	400	river	Rhone	+
L179	12.05.20	46°7'11" 7°4'10"	light rain (3-6°C)	470	inland canal	Canal du Syndicat	+
L180	12.05.20	46°7'7" 7°4'5"	light rain (3-6°C)	460	river	la Drance	-
L181	12.05.20	46°13'12" 7°21'36"	light rain (3-6°C)	480	river	Rhone	+

Water sample ID*	Sampling date (dd.mm.yy)	Location of isolation (DMS)	Weather conditions (temperature)	Altitude (m)	Type of water body	Name of water body	Downstream of WWTP**
L182	12.05.20	46°10'53" 7°25'4"	light rain (3-6°C)	500	river	la Borgne	+
L183	12.05.20	46°16'35" 7°30'30"	light rain (3-6°C)	520	river	Rhone	+
L184	12.05.20	46°18'23" 7°41'33"	light rain (3-6°C)	620	stream	Turtmäna	-
L185	12.05.20	46°18'17" 7°51'21"	cloudy (5-13°C)	640	river	Rhone	+
L186	12.05.20	46°14'43" 7°52'30"	cloudy (5-13°C)	650	river	Vispa	+
L187	12.05.20	46°18'25" 7°56'50"	cloudy (5-13°C)	660	river	Rhone	+
L188	12.05.20	46°23'30" 8°7'32"	cloudy (5-13°C)	990	river	Rhone	+
L189	12.05.20	46°29'11" 8°15'46"	cloudy (5-13°C)	1380	stream	Mistigerbach	-
L190	12.05.20	46°32'10" 8°21'26"	cloudy (5-13°C)	1360	stream	Goneri	-
L191	12.05.20	46°38'32" 8°35'26"	cloudy (5-13°C)	1430	river	Reuss	+

\* Cells highlighted in grey indicate samples that tested positive for *L. monocytogenes*. Strain ID and clonal complex (CC) are indicated in brackets.

\*\* WWTP, wastewater treatment plant; +, WWTP located upstream; -, no WWTP in the vicinity.

**Table S2.** Internalin A amino acid sequence comparison among 25 *Listeria monocytogenes* isolated from surface water

Amino acid position and substitution <sup>1</sup>	Type of amino acid substitution	Strain ID	Remarks
S32N	Non-conservative	L41	CC4 only
V44I	Conservative	L58, L105, L124, L49	
A51T	Non-conservative	L86, L72, L50, L137, L180, L51, L138, L36	
L94V	Conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L72, L50	
D118N	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L72, L50	
S142T	Conservative	L42, L44, L128-3, L28, L174	CC1 only
L157I	Conservative	L86, L137, L180, L51, L138	
S187N	Non-conservative	L52, L58, L105, L124, L49, L72, L50	
A454T	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L50, L72, L36, L137, L180, L51, L138,	
N474S	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L36, L137, L180, L51, L138,	
S476P	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L36, L137, L180, L51, L138,	
A500V	Conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L50, L72, L36, L137, L180, L51, L138, L57, L111, L127, L188	
Y530H	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L36, L137, L180, L51, L138,	
I533V	Conservative	L12, L164, L137, L180, L138, L36, L51	
K539Q	Non-conservative	L137, L180, L138, L51	CC6 only
N544K	Non-conservative	L12, L164	
D558N	Non-conservative	L137, L180, L138, L51, L36, L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52	

Amino acid position and substitution <sup>1</sup>	Type of amino acid substitution	Strain ID	Remarks
L572F	Non-conservative	L12, L164, L137, L180, L138, L36, L51	
E573D	Conservative	L127, L188, L111, L57, L86, L72, L50	
P594A	Non-conservative	L127, L188, L111, L57, L86, L72, L50	
I644V	Conservative	L127, L188, L111, L57, L86	
T648S	Conservative	L36, L137, L180, L51, L138, L86, L57, L111, L127, L188, L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41	
T652A	Non-conservative	L127, L188, L111, L57, L86	
T664A	Non-conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L137, L180, L36	
A671T	Non-conservative	L12, L164	CC6 only
-741N	Non-conservative	L12, L164	CC6 only
-742T	Non-conservative	L12, L164	CC6 only
-743S	Non-conservative	L12, L164	CC6 only
D764E	Conservative	L86, L51, L138	
Y774D	Non-conservative	L58, L105, L124, L49, L12, L164	CC4 and CC6

<sup>1</sup> Amino acid positions are relative to *L. monocytogenes* EGDe InlA protein.

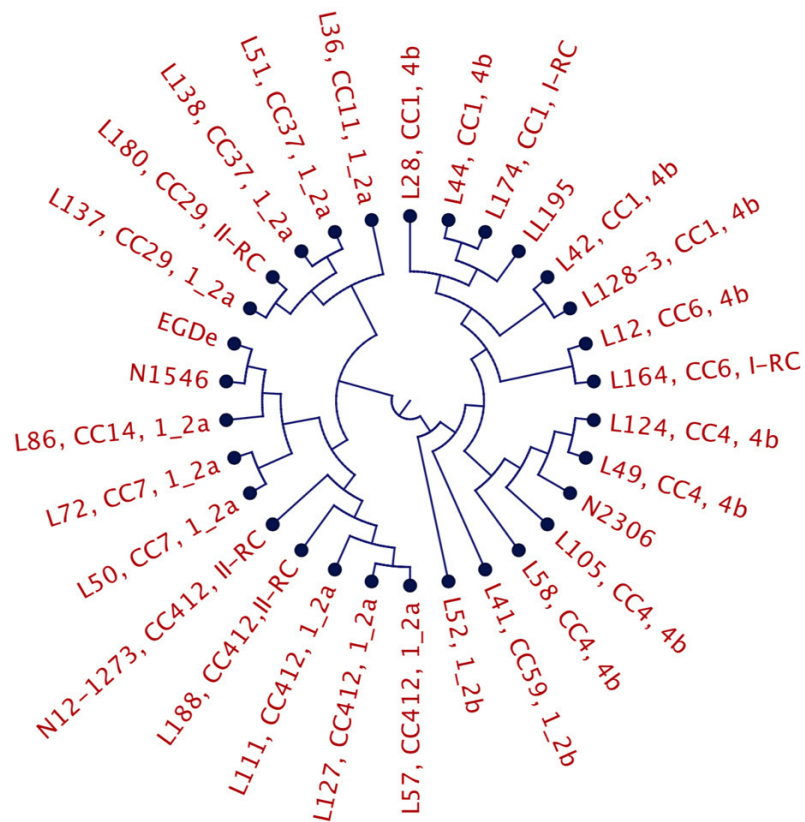
**Table S3:** Presence or absence of virulence factor genes among 25 *Listeria monocytogenes* isolated from surface water

[illegible]

**Table S4:** Listeriolysin O amino acid sequence comparison among 25 *Listeria monocytogenes* isolated from surface water

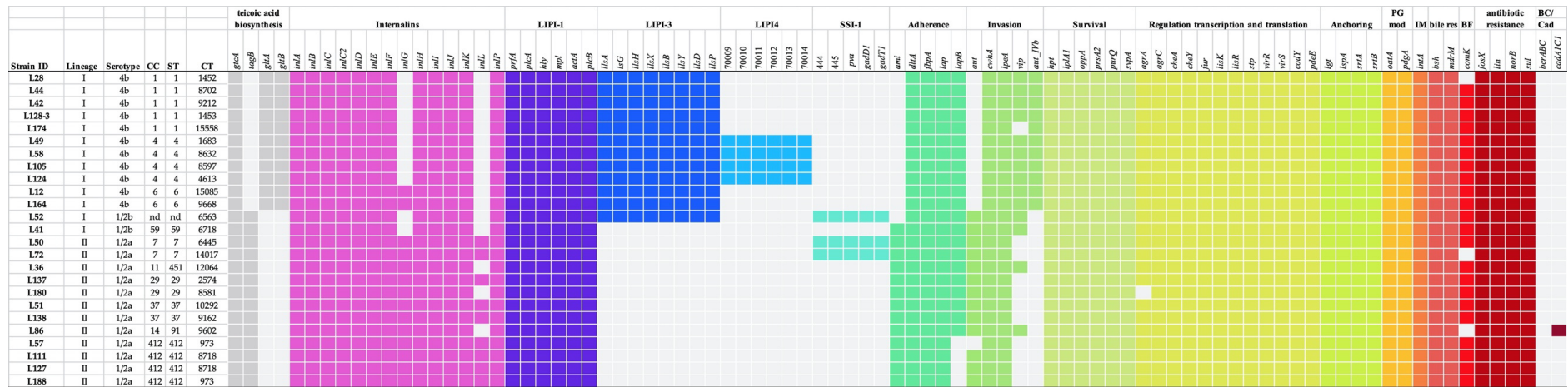
Amino acid position and substitution <sup>1</sup>	Type of amino acid substitution	Strains	Remarks
H31N	Non-conservative	L164,L52,L105, L41,L124,L12,L58, L49	CC412 only
V433I	Conservative	L188,L57,L111,L127	
S523K	Conservative	L28, L44, L42, L128-3, L174, L49, L58, L105, L124, L12, L164, L52, L41, L36	

<sup>1</sup> Amino acid positions are relative to *L. monocytogenes* EGDe LLO protein



**Figure S1:** Phylogeny circular cladogram of the isolates based on average nucleotide identity (ANI) comparison showing the relationship of 25 *Listeria monocytogenes* isolated from surface water and five reference strains.





## **Danksagung/Acknowledgement**

First of all, I thank Prof. Dr. med. vet. Dr. h. c. Roger Stephan for the opportunity to perform this doctoral thesis at the Institute for Food Safety and Hygiene at the University of Zurich. I am especially thankful for his very valuable advices and support throughout the whole time.

My special thank goes to Dr. Katrin Zurfluh for her excellent introduction into my doctoral thesis. She taught me all the laboratory methods I used in my work and was always there to answer my questions in a very professional way.

Furthermore, I would like to thank Dr. Magdalena Nüesch-Inderbinen for her great support in writing the paper.

Last but not least, I would like to thank all employees of the Institute of Food Safety and Hygiene (ILS) for the cooperativeness and goodwill.

I am grateful for the great support of my family, especially Sybille and Cosimo.

# Curriculum Vitae

Vorname Name	Susanne Raschle
Geburtsdatum	27.05.1988
Geburtsort	Münsterlingen TG
Nationalität	Schweizerin
Heimatort	Mosnang SG

## Schul Ausbildung

Aug/1995 – Jun/2001	Primarschule, Kreuzlingen, Schweiz
Aug/2001 – Jun/2004	Sekundarschule, Kreuzlingen, Schweiz
Aug/2004 – Jun/2007	Fachmittelschule, Romanshorn, Schweiz

## Höchster Schulabschluss

Jun/2007	Fachmittelschulabschluss
----------	--------------------------

## Studium

Sep/2007 – Jun/2011	Zürcher Fachhochschule für Angewandte Wissenschaften, Winterthur, Schweiz
Jun/2011	Bachelor of Sciences ZFH in Physiotherapie, Winterthur, Schweiz
Sept/2014 – Jul/2019	Bachelor/Master of Veterinary Medicine, Universität Zürich, Zürich, Schweiz

## Abschlussprüfung vet. med.

Dez/2019	Universität Zürich, Zürich, Schweiz
----------	-------------------------------------

Jan/2020 – Jun/2020

## Anfertigung der Dissertation

unter Leitung von Roger Stephan  
am Institut für Lebensmittelsicherheit und -hygiene  
der Vetsuisse-Fakultät Universität Zürich  
Direktor Roger Stephan

**Anstellung nach Abschluss des veterinärmedizinischen  
Studiums**

Jan/2020-Jun/2020

Doktorandin Institut für Lebensmittelsicherheit und -  
hygiene, Vetsuisse Zürich

Seit Jul/2020

Assistentztierärztin, Tierarztpraxis, Berg, Schweiz